

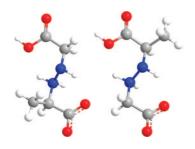
Structure of N,N'-Bis(amino acids) in the Solid State and in Solution. A ¹³C and ¹⁵N CPMAS NMR Study

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Three bis(amino acids) linked by the amino groups have been prepared and structurally characterized. We have named them Gly-Gly, Ala-Ala and Gly-Ala (or Ala-Gly). These compounds have been characterized by NMR both in solution and in the solid state. They exist as zwitterions with the ammonium group proximal to the carboxylate anion. In the case of Gly-Ala, a dynamic situation is observed by CPMAS NMR (¹³C and ¹⁵N) corresponding to a double proton migration between two proximal tautomers.

 α -Amino acids and their corresponding peptides have been modified in any imaginable way: conformationally restricted, pseudo, non-natural, C,C'-linked, etc.\(^1\) The peptide bond in I has been modified to yield two well-known derivatives having

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a N-N bond: the hydrazinopeptides \mathbf{H}^2 and the *N*-amino peptides \mathbf{HI} .

Surprisingly, a third possibility (IV) has almost never been explored (no compound with the connectivity present in **IV** is reported in the CSD).⁴ We describe here the synthesis of the three simplest N,N'-bis(amino acids) and their structural study by NMR supported by theoretical calculations. Scheme 1 presents the compounds 1-5 studied in this work, including the three N.N'-bis(amino acids) Gly-Gly, Ala-Ala, and Gly-Ala (or Ala-Gly). It should be noted that in the bis(amino acids), Ala stands for racemic alanine. In the case of the Ala-Ala derivative, only the meso compound was synthesized and studied. Besides the structures of Scheme 1, we have calculated the neutral forms [H₂N-CH(R)-CO₂H, HO₂C-CH(R)-NH-NH-CH(R)-CO₂H] as well as the double zwitterions [-O₂C-CH(R)-NH₂⁺-NH₂⁺-CH(R)-CO₂⁻] of Gly-Gly, Ala-Ala and Ala-Gly, but since they are not observed, we report the results in Supporting Information.

More than 100 years ago, Thiele and Bailey⁵ prepared Ala-Ala as a mixture of diastereoisomers through an intermediate hydrazinopropionic acid derivative. The unsubstituted counterpart Gly-Gly was synthesized by Gisin and Brenner⁶ starting from glyoxylic acid and later by Arakawa et al.⁷ through a hetero Diels—Alder reaction. Gly-Gly has important applications in photography and ~50 of Fuji's patents deal with this compound.^{8,9} To our knowledge, the preparation of Ala-Gly has not been reported before. In the present work, the synthesis of the bis(amino acids) Gly-Gly (3), Ala-Ala (4), and Ala-Gly (5) was carried out following a common route resulting from modification of the procedure described by Arakawa et al.⁷ for 3. After condensation of di-*tert*-butyl azodicarboxylate with the corre-

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SCHEME 1. Molecules under Study

SCHEME 2. Synthetic Procedure

sponding diene (Scheme 2), the C=C double bond in the resulting adduct (6–8) was subjected to oxidative cleavage by treatment with ruthenium(IV) oxide to yield the diacids 9–11. Removal of the Boc protecting groups in 3 N HCl at room temperature followed by treatment with an ion-exchange resin afforded the desired compounds 3–5 in 60–88% global yield. In the original protocol, the diacid 9 was transformed into the methyl diester prior to hydrolysis. We found no advantage in carrying out this additional step. On the contrary, hydrolysis of the diester required harsher conditions (heating in 6 N HCl), which could result in cleavage of the N–N bond.

We have gathered in Table 1 the NMR results concerning compounds **1–5** and represented in Figure 1 the ¹³C and ¹⁵N CPMAS spectra corresponding to Gly-Gly, Ala-Ala, and Ala-Gly (or Gly-Ala).

All structures represented in Scheme 1 have been calculated at the B3LYP/6-31G* level^{12,13} using the solvation model of

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TABLE 1. NMR Results (δ ppm)

TABLE 1. INTR Results (0 ppm)									
compd/conditions	CH(R)	CH_3	СО	N	ref				
1 Gly									
water neutral	42.8		173.6	-347.3	13 C, 10 15 N 11				
CPMAS	43.3		176.1	-344.8	this work				
1H ⁺ GlyH ⁺									
water acid	41.5		171.2	-345.4	13 C, 10 15 N 11				
1 ⁻ Gly ⁻					40 44				
water basic	46.0		182.7	-354.7	13 C, 10 15 N 11				
2 Ala									
water neutral	51.9	17.5	177.0	-331.1	$^{13}\text{C},^{10}$ $^{15}\text{N}^{11}$				
CPMAS	50.5	20.0	177.4	-334.6	this work				
2H ⁺ AlaH ⁺					12 - 10 15 -11				
water acid	50.1	16.5	174.0	-331.3	$^{13}\text{C},^{10}$ $^{15}\text{N}^{11}$				
2- Ala-					12 ~10				
water basic	52.7	21.7	185.7		$^{13}C^{10}$				
3 Gly-Gly	51.5		172.0	2066	.1.1				
CD ₃ OD ^a	51.7		173.0	-306.6	this work				
CPMAS	50.0		177.8	-292.3	this work				
4 Ala-Ala	54.0		173.6	-305.6					
CD_3OD^b	59.2°	15.9^{d}	176.0	-297.3	this work				
CD3OD CPMAS	55.3	17.7	170.0	-297.5 -281.5	this work				
CFMAS	61.0	16.7	176.6	-281.5 -282.6	uiis work				
	01.0	15.1	175.7	-282.0 -292.0					
		13.1	1/3./	-293.8					
5 Gly-Ala				273.0					
CD_3OD^e	$51.5^{f,g}$		172.6^{g}	-308.1	this work				
CD3OD	$58.8^{h,i}$	15.4^{d}	175.0^{i}	-296.4	tins work				
CD ₃ CN ^j	52.1^{g}	10	173.9^g	not obsd	this work				
,01,	58.9^{i}	16.1	176.3^{i}	not obsd					
$CPMAS^k$	51.1 ^g		175.1	-297	this work				
	57.9^{i}	15.3							

 a <3.6 mg/0.75 mL. b 4 mg/0.75 mL. c 1J = 144.7 Hz. d 1J = 129.0 Hz. e <4.3 mg/0.75 mL. f 1J = 140.3 Hz. g Gly. h 1J = 143.8 Hz. i Ala. j <5.6 mg/0.75 mL. k See spectra (Figure 1).

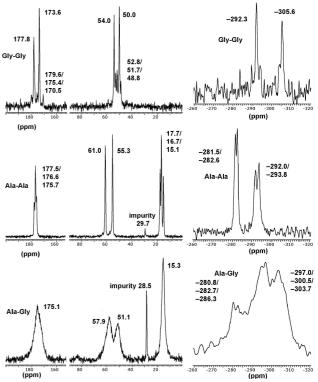


FIGURE 1. ¹³C (left) and ¹⁵N (right) CPMAS NMR spectra recorded at 100.73 and 40.60 MHz, respectively.

Tomasi (PCM) with water as solvent and Pauling's van der Waals radii. 14 Without solvent, in all cases the only minima

correspond to neutral molecules (no proton transfer between the carboxylic acid and the amino group occurs). Over the optimized geometries, absolute shieldings σ were calculated with the GIAO approximation. ¹⁵

Concerning the energies including solvation, the following values have been obtained: Gly-Gly, distal 3.18 kJ mol^{-1} higher than proximal; Ala-Ala, distal 8.49 kJ mol^{-1} higher than proximal. Thus, although proximal zwitterions are always preferred, the difference is higher for N,N'-bis(alanine) than for N,N'-bis(glycine). In the case of the N-glycine-N'-alanine derivative, the most stable tautomer is the Ala-Gly $\mathbf{5a}$, followed by the Gly-Ala $\mathbf{5b}$ ($+0.89 \text{ kJ mol}^{-1}$) [at 25 °C, this corresponds to 59% of $\mathbf{5a}$ and 41% of $\mathbf{5b}$]. Considering the three equilibria, this may indicate that the neutral -NH-CH(R)-CO₂H group is more stable for $R = CH_3$ than for R = H. The distal tautomers are less stable, $\mathbf{5c}$ +3.65 [at 25 °C, this corresponds to 82% of $\mathbf{5a}$ and 18% of $\mathbf{5c}$] and $\mathbf{5d}$ +5.58 kJ mol⁻¹.

The σ values are given in Supporting Information. In Table 2 are reported the corresponding chemical shifts. To transform the absolute shieldings into chemical shifts in solution (CD₃OD and CD₃CN) and in the solid state (CPMAS), the four following equations have been used:

Solution:
$$\delta^{13}$$
C = 203.3 - 1.094 σ^{13} C, n = 20, r^2 = 1.000
Solution: δ^{15} N = -188.9 - 0.69 σ^{15} N, n = 7, r^2 = 0.971
Solid: δ^{13} C = 204.5 - 1.098 σ^{13} C, n = 15, r^2 = 1.999
Solid: δ^{15} N = -150.8 - 0.876 σ^{15} N, n = 6, r^2 = 0.992

The average values were calculated for 50:50 mixtures in the cases of Gly-Gly and Ala-Ala, i.e., assuming a rapid equilibrium between two identical tautomers, 3a/3a, 3b/3b, 4a/4a, or 4b/4b. In the case of the mixed *N*,*N*'-bis(amino acid), we have used the theoretically calculated 59:41 mixture.

It is well-known that protonation effects on the amino and hydrazino groups are weak or very weak both in ¹³C and in ¹⁵N NMR. ¹⁶ Therefore, no great differences are expected among the different structures of the *N*,*N*′-bis(amino acids) under study.

In solution and using the 13 C chemical shifts, it is difficult to assign the structure to the proximal (fitted values) or to the distal (predicted values) situations because in both cases the correlation coefficient is $r^2 = 1.000$. However, the intercept is closer to 0 (0.6 vs 2.4) and the slope is closer to 1 (0.99 vs 0.97) for the proximal structure than for the distal structure. The situation becomes clearer using the 15 N chemical shifts (adding the five data of 1, 1H⁺, 1⁻, 2, and 2H⁺). For the proximal, the intercept

TABLE 2. Calculated NMR Chemical Shifts^a

TABLE 2. Calculated NMR Chemical Shifts"								
compd/phase ^b	near	CH(R)	CH_3	CO	N			
1 Gly								
solution		41.9		173.4	-342.2			
solid		42.5		174.5	-345.3			
1H ⁺ GlyH ⁺								
solution		42.7		170.2	-345.6			
1- Gly-								
solution		45.8		184.3	-352.4			
2 Ala								
solution		52.8	15.7	176.4	-334.2			
solid		53.3	16.2	177.5	-335.1			
2H ⁺ AlaH ⁺								
solution		53.6	16.7	172.9	-337.4			
2 ⁻ Ala ⁻								
solution		51.2	21.6	185.0	-348.2			
3a Gly-Gly prox								
solution	average ^c	50.8		174.6	-304.8			
solid	CO_2H	51.4		175.6	-301.5^d			
	CO_2^-	53.3		173.4	-294.2^{e}			
3b Gly-Gly dist								
solution	average ^c	50.0		175.5	-307.5			
solid	CO_2H	51.6		171.8	-312.1^{d}			
	CO_2^-	49.7		180.9	-290.4^{e}			
4a Ala-Ala prox								
solution	average ^c	61.4	14.4	175.6	-297.1			
solid	CO_2H	58.6	15.2	177.3	-291.9^d			
	CO_2^-	65.3	14.4	175.9	-284.2^{e}			
4b Ala-Ala dist								
solution	average ^c	59.6	13.6	178.3	-303.1			
solid	CO_2H	62.9	13.9	174.3	-312.1^d			
	CO_2^-	57.6	14.5	183.9	-290.4^{e}			
5a/5b	f CI	51.7		172.5	207.4			
solution	av ^f Gly	51.7	12.5	173.5	-307.4			
7. A1. C1	av ^f Ala	60.1	13.5	176.1	-295.4			
5a Ala-Gly prox	CI	52.2		172 (201.1			
solid	Gly	53.3	140	173.6	-301.1			
51. Cl., A1.,	Ala	58.9	14.8	177.7	-286.0			
5b Gly-Ala prox	Clv	510		175 7	202.0			
solid	Gly	51.0	12.6	175.7	-303.0			
Fo Alo Clyr dist	Ala	63.2	12.6	176.5	-294.1			
5c Ala-Gly dist solid	Clv	49.5		180.9	-289.3			
Solid	Gly Ala	63.2	13.6	180.9 174.1	-269.5 -303.5			
Ed Cly Ale dist	Ala	03.2	15.0	1/4.1	-303.3			
5d Gly-Ala dist solid	Gly	51.7		171.7	-319.1			
Solid	Ala	57.6	14.4	184.2	-319.1 -281.7			
5a/5b	Ala	37.0	14.4	104.2	-201.7			
solid	av ^c Gly	52.2		174.6	-302.0			
SOHU	av Gly av ^c Ala	61.0	13.7	174.0 177.1	-302.0 -290.0			
5c/5d	av Aid	01.0	13./	1//.1	270.0			
solid	av ^c Gly	50.6		176.3	-304.2			
50110	av Gly av ^c Ala	60.4	14.0	170.3 179.1	-292.6			
	av Ala	00.7	17.0	1//.1	272.0			

 $[^]a\,\delta$ ppm; roman = fitted; italic = predicted. b "Solution" means the average situation found in methanol or acetonitrile, and "solid" represents the unique tautomer found in the solid state with the exception of compound 5. c 50:50. d NH₂+. e NH. f 59:41.

is 0 and the slope is 1, whereas for the distal, the intercept is 38.4 and the slope is 1.1.

For the proximal Ala-Gly (Gly-Ala) derivative, 5a/5b, in CD₃OD the calculations predict values very close to the experimental ones: experimental (Table 1)/calculated (Table 2): 51.5/51.7, 172.6/173.5, -308.1/-307.4, 58.8/60.1, 15.4/13.5, 175.0/176.1 and -296.4/-295.4 ppm.

In the solid state, there are two different cases: Gly-Gly and Ala-Ala present narrow signals in both ¹³C and ¹⁵N CPMAS NMR spectra. The calculations (Table 2) reproduce well the positions of all signals, but mainly in Ala-Ala there is a clear splitting of several signals, e.g., the methyl group (three signals instead of two), the carboxylic group (three signals instead of

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SCHEME 3. Possible Solid-State Intramolecular Double Proton Transfers

two), and both nitrogen atoms (two signals instead of one). Since the distal isomer **4b** can be excluded (no signal at -312 ppm in 15 N NMR), the multiplicity must be related to the existence of two independent molecules in the unit cell, probably two different conformers. In the Gly-Gly derivative, there are some small signals in the 13 C NMR spectrum (Figure 1) that could correspond to a minor conformer.

The Ala-Gly (Gly-Ala) derivative behaves differently. In both spectra (Figure 1) the signals are broad (the narrowness of the impurity at 28.5 ppm proves that this is not due to a lack of resolution). We assign this broadening to a rapid equilibrium between **5a** and **5b** that we assumed are present in a 50:50 mixture in the solid state (Scheme 3). The calculated average signals (Table 2) agree reasonably well with the broad signals: 13.7/15.3, 52.2/51.1, 61.0/57.9, 174.6–177.1/175.1 ppm (13 C) and -290 and -302 ppm with the large signal between -281 and -304 ppm (Figure 1, 15 N). Note that this signal can be described as the sum of two signals at -282.7 and -303.7 ppm (slow proton transfer) plus a central signal at -297.0 ppm (fast proton exchange, calculated at -296.0 ppm).

The other possibility, a rapid equilibrium in the solid state between two Ala-Gly tautomers **5a** and **5c** should be rejected for the two following reasons. Solid state proton transfer (SSPT) occurs only between tautomers of identical or very similar energies;¹⁷ the difference being 0.89 kJ mol⁻¹ for the **5a/5b** pair but 3.65 kJ mol⁻¹ for the **5a/5c** pair. Another reason is that 1,2-proton transfers in hydrazine derivatives (**5a/5c** pair) are generally forbidden.¹⁸

In summary, the study of the molecules described in this paper has resulted in the observation of interesting solid state properties, including a new example of the rare phenomenon of SSPT. Further studies will be needed to finish this work, including X-ray structure determinations (for the moment, it has been impossible to grow convenient crystals) and ¹⁵N-labeled molecules for CPMAS low-temperature NMR studies.

Experimental Section

Characterization of compounds 3-11 is given in Supporting Information.

General Procedure for the Hetero Diels—Alder Reaction. To a solution of di-*tert*-butyl azodicarboxylate (4.0 g, 17.39 mmol) in CCl_4 (40 mL) cooled at -40 °C was added the corresponding diene (1.5–5.0 equiv) dissolved in CCl_4 (20 mL). The reaction mixture was stirred at this temperature for 2 h and then at room temperature for 7 days. After removal of the solvent, the remaining residue was purified by column chromatography on silica gel (eluent, hexanes/ EtOAc 9:1) to afford 6-8.

General Procedure for Double Bond Cleavage. A solution of 6-8 (7.74 mmol) in EtOAc (40 mL) was cooled to 0 °C, and a 10% aqueous solution of sodium periodate (70 mL) containing ruthenium(IV) oxide (30 mg, 0.23 mmol) was added. The resulting biphasic system was vigorously stirred at 0 °C for 2 h and then at room temperature overnight. The organic layer was separated, and the remaining aqueous layer was extracted with EtOAc (3 × 30 mL). The combined organic extracts were dried over anhydrous MgSO₄ and filtered. Evaporation of the solvent yielded a black oil, which was purified by column chromatography on silica gel (eluent, Et₂O) to afford 9-11.

General Procedure for *N*-Boc Deprotection and Isolation of the Final Bis(amino acid). A 3 N solution of HCl in anhydrous EtOAc (5 mL) was added to 9–11 (1.67 mmol), and the reaction mixture was stirred at room temperature for 1 h. After evaporation of the solvent, the residue was taken up in water and lyophilized. It was then redissolved in water (10 mL) and Amberlyst A26 (hydroxyde form) (950 mg, wet wt) was added. The mixture was stirred manually occasionally until the resin color completely turned from pink to pale yellow (about 2 h). The resin was removed by filtration and washed thoroughly with water. Concentration of the aqueous solution followed by lyophilization afforded 3–5.

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Supporting Information Available: ¹H and ¹³C NMR spectra and characterization of all compounds synthesized. Details on NMR structural studies and theoretical calculations. Total energy, nuclear shieldings and Cartesian coordinates of the molecules calculated at the B3LYP/6-31G(d,p) computational level. This material is available free of charge via the Internet at http://pubs.acs.org.

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